Preparation

HCL rinse Lab 201

1. Set up catchment bin in sink, with a container with baking soda within it for waste HCL.
2. Rinse 8” dissecting dish with 1 M HCL and dump into waste HCL container. Set aside on the counter.
3. Fill first vial about halfway with HCL, cap and shake a few times
4. Remove cap, dump HCL into next vial and set in the dissecting dish. Cap the new vial, shake a few times
5. Repeat step 4 until all vials are rinsed with HCL
6. Pour remaining HCL in waste container
7. Rinse each vial and lid in DI water and dump waste water into the catchment bin
8. Check pH of catchment bin and waste HCL, add baking soda until neutralized. Rinse down the drain with lots of water.
9. Place lids in previously cleaned beaker
10. Place dissecting dish with vials in drying oven (lab 102)at 50C for at least 2 hrs

Small eye delamination

1. Take batch of eyes out of freezer and place in fridge night before (Wednesday’s eyes were done this way)
   1. Note: small eyes will also thaw quickly in a DI water bath, so can take them out just prior to dissection
2. Prepare workstation
   1. Collect DI water in clean beaker and/or squirt bottle. Draw fresh DI each day from the nano pure in Lab 201
   2. Clean tools and dissecting dish by wiping with ETOH wetted chemwipe
   3. Blast with compressed air to remove any particles
   4. Allow to dry
   5. Get new scalpel blade ready
3. Label vials with species, sample #, Left or right, and layer number.
   1. Note: for small eyes, there are generally just an “N” layer for nucleus and “R” layer for remainder. If the nucleus breaks up, it can be split into more than one vial.
   2. Record each layer with ID on the vial data sheet
   3. Weigh empty vials without lids on scale in lab 102, record on data sheet “vial\_MT\_wt”
4. Remove eye from baggie and place in large dissecting dish with label
5. Take picture of the eye with the label
6. Use fresh scalpel to dissect lens from globe of the eye and place eye in smaller glass dish (see video)
7. Clean tools with etoh wetted wipe and blast with compressed air
8. Gently dissect outer lens from core (see video), use dissecting scope if necessary
   1. If nucleus falls apart attempt to keep remainder, nucleus parts and nucleus core separated
9. Place nucleus core in small beaker and rinse with a few drops of DI water, remove from DI water and place in appropriate vial
10. Repeat step 9 with the next layer out, until all layers are in vials
11. Place in fridge until going into drying oven in lab 102
12. Clean tools for next dissection